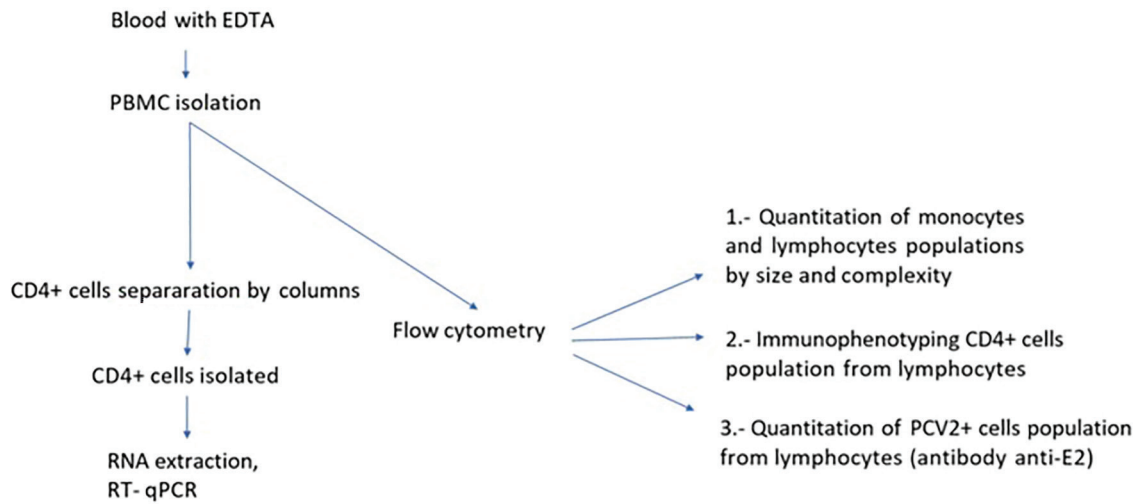


Supplementary material

Diagram

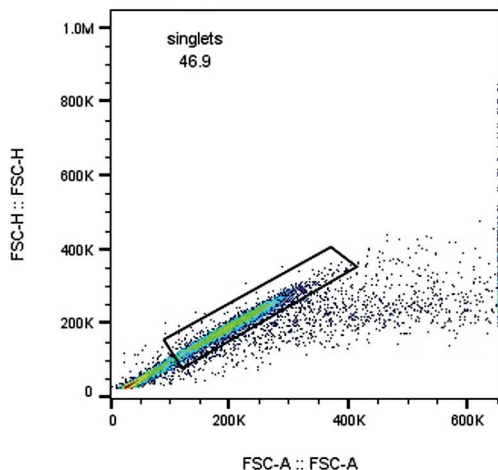


***The same procedure was performed for control and infected groups**

- First, the gating was based on forward scatter (size) and side scatter (cell complexity) to select cells showing homogeneous size and complexity as indicator of lymphocytes.
- Second, we standardized the gating parameters for CD4+ cells. PBMCs were selected with an anti-CD4 antibody and dot plots. Histograms were used to set gating parameters.
- Third, from the gating strategy generated in the two, previous, separate experiments we proceeded to separate cells from all the samples and in all the experiments.

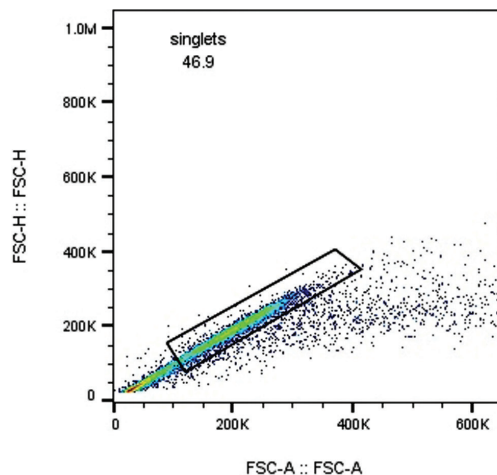
1. Quantitation of monocyte and lymphocyte populations by size and complexity

Singlets from PBMC



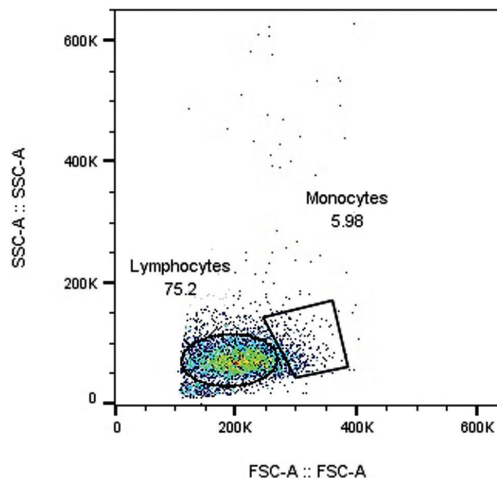
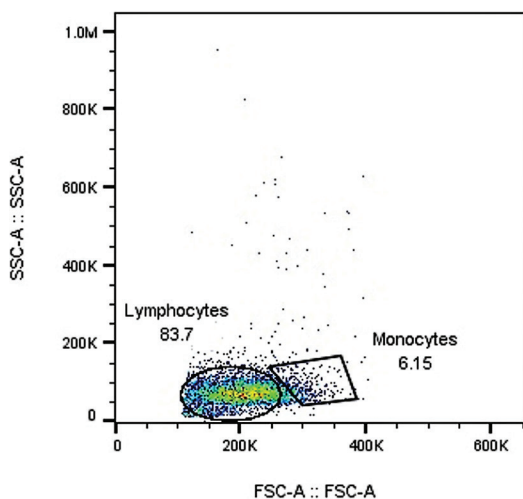
2. Immunophenotyping CD4+ cells population from lymphocytes

Singlets cells from PBMC



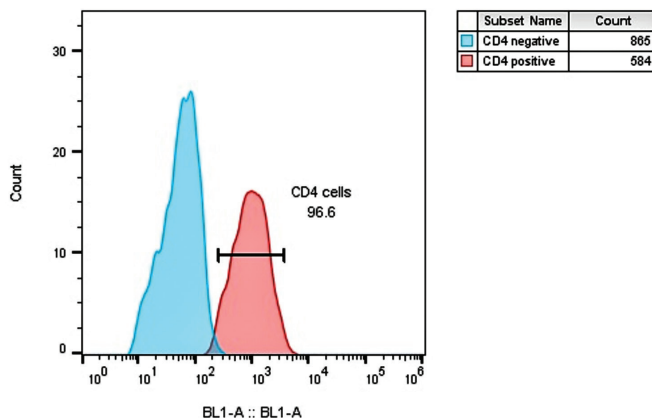
Lymphocytes and monocytes populations from singlets were analyzed according to their size and complexity.

Lymphocytes from singlets for CD4+ cell immunodetection



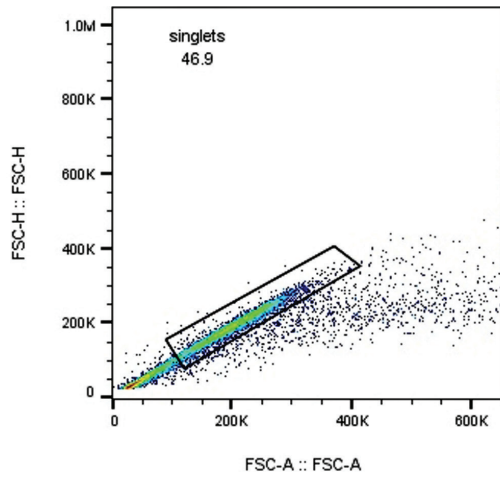
Separation according to: University of Kansas Medical Center. Online-protocol: <https://www.kumc.edu/Documents/flow/Peripheral%20Blood%20Mononuclear%20Cell%20and%20RBC%20lysis.pdf>

Analysis of negative cells versus CD4+ cells.

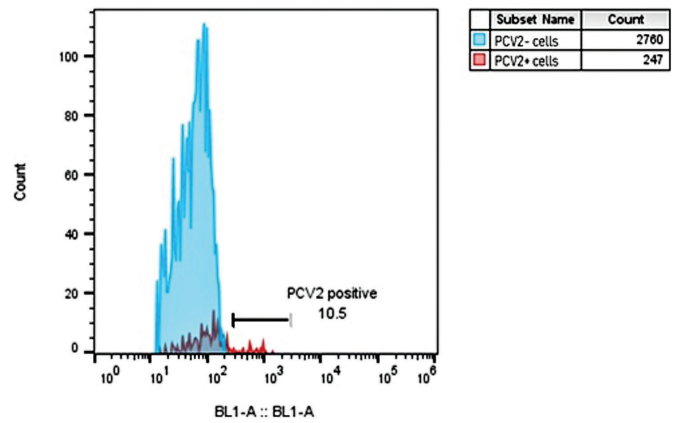


3. Quantitation of PCV-2+ cells population from lymphocytes (antibody anti-E2)

Singlets from PBMC



Analysis of PCV-2- cells *versus* PCV-2+ cells population.



Lymphocytes from singlets for PCV+ cell immunodetection with antibodies against E2 protein.

